

UNIVERSITI TEKNOLOGI MARA

**ANTIPROLIFERATIVE EFFECT OF
CASSIA AURICULATA
(CAESALPINIACEAE) FLOWERS**

SAMARA YAHYA KRAIDI

Thesis submitted in partial fulfillment of the
requirements for the degree of
Master of Science

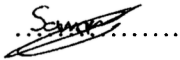
Faculty of Pharmacy

August 2014

AUTHOR'S DECLARATION

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Name of Student : Samara Yahya Kraid
Student I.D. NO. : 2010832302
Programme : Master of Science (PH780)
Faculty : Faculty of Pharmacy
Thesis Title : Antiproliferative effect of *Cassia auriculata* flowers
Signature of Student : 
Date : August 2014

ABSTRACT

Cassia auriculata is a common South Asian medicinal plant which is widely used in traditional medicine. The plant has been used in folk medicine to treat various disease conditions and the pharmacological activity of different parts of the plant has been well documented as anti-diabetic, anti-oxidant and anti-hyperlipidemic. This study was aimed at investigating anti-proliferative activity of *C. auriculata* flowers. Cellular viability was measured by (MTS) assay. Effects on apoptotic (programed cell death) machinery were examined using annexin-V and propidium iodide staining using flow cytometry. Molecular mechanisms were analyzed by real-time polymerase chain reaction (RT-PCR). Effects on cell cycle were determined by cell cycle analysis using flow cytometry. Crude extract of *C. auriculata* flowers was prepared by extraction with a mixture of methanol and dichloromethane. The results showed that crude extract exhibited specific anti-proliferative effect against liver cancer (HepG2) cell line with IC_{50} value of 63 ± 2.2 $\mu\text{g/ml}$ as compared to effects on mammary cancer (MCF-7) and colon cancer (HCT116) cell lines with IC_{50} values of 125 ± 2.6 and 199 ± 5.8 $\mu\text{g/ml}$, respectively. Crude extract was selective to liver cancer cells as it showed minimal cytotoxic effect against normal embryonic liver (WRL-68) cell line with IC_{50} of 251 ± 4.1 $\mu\text{g/ml}$. Partitioning and fractionation of crude extract yielded six fractions of which, fraction1 elicited most potent anti-proliferative effect in HepG2 cells with an IC_{50} of 12.5 ± 2.3 $\mu\text{g/ml}$. Mechanism of cell death was via apoptosis which was significantly induced in a dose-dependent manner in HepG2 cells treated with crude extract or with fraction1 at three different concentrations which corresponded to their inhibitory concentrations in HepG2 cells (IC_{20} , IC_{50} and IC_{70}). By comparison, fraction1 was more potent than crude extract at induction of apoptosis. Induction of apoptosis by both crude extract and fraction1 occurred via up-regulation in expression levels of p53, a tumor suppressor gene; Bax, a pro apoptotic gene; caspase-3, a major apoptotic gene and simultaneous down-regulation of Bcl-2, an anti-apoptotic gene. Cell cycle analysis showed crude extract and fraction1 elicited arrest of cells at G_2/M phase. Treatment of HepG2 with F1 caused marked accumulation of cells at G_2/M phase and a corresponding decrease in cells at G_0/G_1 and S phases. These results show that *C. auriculata* flower extract and its purified fraction exerted anti-proliferative effect in HepG2 cells via induction of apoptosis and arrest of the cell cycle at G_2/M phase. There is potential to further develop *C. auriculata* flowers for adjuvant management of hepatocellular carcinoma. More work is required to examine the phytochemistry of *C. auriculata* in relation to its anti-proliferative effect in liver cancer cells.

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